Characterization of the physical and microbiological properties of a powder excipient base (Loxasperse)

Banov, D., Vieira-Banov, F., Carvalho, M. and De Almeida, M. PCCA, 9901 South Wilcrest Drive, Houston, TX 77099, USA

Loxasperse is a proprietary blend of micronized xylitol and micronized poloxamers, designed to be mixed with active substances in order to improve their water solubility and dispersibility. The use of xylitol and poloxamers in nebulization and irrigation is thoroughly referenced in the literature and there is evidence-base for their safety and efficacy (Durairaj et al., 2007; Plataki et al., 2011). Xylitol and poloxamers exhibit antimicrobial activity and, therefore, Loxasperse was also designed to prevent microbial growth (Veyries et al., 2000; Zabner et al., 2000). Loxasperse mixtures are thus dry powders, formulated as non-sterile capsules or sachets, indicated to be dissolved in purified water or saline prior to the administration of compounded medicines for nebulization and irrigation (Figure 1).



Methodology



Physical properties: To determine the particle size distribution of Loxasperse and Loxasperse with

Figure 1 Powder excipient base (Loxasperse)

itraconazole, two different tests were performed, respectively: static laser light scattering and optical microscopy. Microbiological properties: To characterize the antimicrobial activity of Loxasperse with itraconazole, two different Minimum inhibitory Concentration (MIC) methods were performed against a suite of fungal and bacterial strains, as follows: broth microdilution method and agar dilution method. All strains were obtained from the American Type Culture Collection (ATCC). To estimate the microbiological growth in Loxasperse, the water activity of the powder excipient base was determined using the AquaLab Water Activity Meter (AquaLab, 2008; 2013).

Results & Discussion



Particle size distribution: The particles in a sample are not perfectly mono-disperse (i.e. every single particle has exactly the same dimensions) but, instead, these commonly consist of a statistical distribution of particles of different dimensions. Several tests may be performed in order to characterize this physical property (Malvern, 2012).

Static laser light scattering: This test provides a volume weighed distribution, in which the contribution of each particle in the distribution relates to the volume of that particle (Malvern, 2012). Loxasperse 6.4% in purified water exhibits a narrow distribution of particles (Figure 2), which demonstrates the optimal physical characteristics and performance of the powder excipient base.

Optical microscopy: The microscopy examination is suitable to determine the distribution of particles of inhalable size (European Commission JRC, 2002) and, therefore, optical microscopy was performed to characterize the effect of Loxasperse in the particle size distribution of itraconazole. An AmScope Microscope Digital Camera was used for photographic characterization of itraconazole 1% in purified water, with and without Loxasperse, at 400x magnification (AmScope, 2013). This test was performed in accordance with the respective 'Physical Test' of the USP Pharmacopoea (The United States Pharmacopeial Convention, 2013). It was observed that, following the addition of Loxasperse, large aggregates of itraconazole were converted in small aggregates and single particles (Figure 3). It is therefore concluded that Loxasperse optimizes the particle size distribution of itraconazole in purified water. Minimum inhibitory concentration: The MIC is the lowest concentration of an antimicrobial that will inhibit the visible growth of a microorganism after overnight incubation. The MIC is the gold standard research tool to determine the in vitro activity of antimicrobials (Andrews, 2001). A lower MIC is indicative of a better antimicrobial agent.

Broth microdilution method: The *in vitro* antifungal activity of itraconazole and Loxasperse with itraconazole (9:1) was determined against 4 fungal strains using the National Committee for Clinical Laboratory Standards (NCCLS) reference methods for yeast and filamentous fungi (Espinel-Ingroff, 2002; NCCLS, 2002a; 2002b). A lower MIC was found in Loxasperse with itraconazole than in itraconazol itself (Table 1). It is concluded that the Loxasperse mixture has improved antifungal activity against all fungal strains tested.

Agar dilution method: The in vitro antimicrobial activity of Loxasperse was determined against 7 microbial strains. A MIC of 17% of Loxasperse was achieved for the majority of the microbial strains tested (Table 2). No antimicrobials were added. Water activity: The a_w is defined as the amount of free or available water in a system and it is a measure of how efficiently the water can take part in a chemical reaction. Reducing the a_w minimizes undesirable chemical reactions and microbiological growth. Most bacteria do not grow at a_w <0.91 and no microbiological growth is possible at a_w <0.60. The a_w is a better index for microbial growth than the water content (Blandamer et al., 2005; AquaLab, 2008; 2013). The aw of Loxasperse was determined after 90 days of storage at three different temperatures, with and without desiccant, and an average of a_w =0.321 and a_w =0.456 was obtained (Table 3). It is concluded that no microbiological growth is possible in Loxasperse, after 90 days of storage at T<45°C, because of its low a_w (<0.60).

Loxasperse with itraconazole has improved particle size distribution in water and also improved antifungal activity in comparison to itraconazole itself. It is concluded that Loxasperse improves the water dispersibility and antifungal activity of itraconazole. Considering the MIC and a_w of Loxasperse alone, it is also concluded that, as expected, Loxasperse prevents microbial growth.

References



Andrews, J. (2001) 'Determination of minimum inhibitory concentrations', Journal of Antimicrobial Chemotherapy, 48 (S1), p.5-16.

AquaLab (2008) Water Activity Meter Operator's Manual. Washington: Decagon Devices, Inc.

AquaLab (2013) Microbial Growth. Available at: http://www.aqualab.com/applications/microbial-growth (Accessed: 10 May 2013).

AmScope (2013) Microscope Cameras. Available at: http://www.amscope.com/Camera.html (Accessed: 8 June 2013).

Blandamer, M., Engberts, J., Gleeson, P. and Reis, J. (2005) 'Activity of water in aqueous systems: A frequently neglected property', Chemical Society Reviews, 34, p.440-58.

Durairaj, L., Launspach, J., Watt, J.L., Mohamad, Z., Kline, J. and Zabner, J. (2007) 'Safety assessment of inhaled xylitol in subjects with cystic fibrosis', Journal of Cystic Fibrosis, 6 (1), p.31-4.

Espinel-Ingroff, A. (2002) 'Antifungal susceptibility methods and their potential clinical relevance', LabMedicine, 8 (33), p.626-31.

European Commission JRC (2002) Guidance Document on the Determination of Particle Size Distribution, Fibre Length and Diameter Distribution of Chemical Substances [Online]. Available at: http://publications.jrc.ec.europa.eu/repository/bitstream/111111111/5555/1/EUR%2020268%20EN.pdf (Accessed: 8 June 2013).

Malvern (2012) 'A Basic Guide to Particle Characterization', *Inform White Paper*. Worcestershire: Malvern Instruments Limited.

Martindale 35 (2007) [CD-ROOM]. RPSGB and Sweetman, S. (ed.). Available: Pharmaceutical Press.

NCCLS (2002a) Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Approved Standard. 2nd edn. Pennsylvania: NCCLS. NCCLS (2002b) Reference Method for Broth Dilution Antifungal Susceptibility Testing of Filamentous Fungi; Approved Standard. Pennsylvania: NCCLS. Plataki, M., Lee, Y., Rasmussen, D. and Hubmayr, R. (2011) 'Poloxamer 188 facilitates the repair of alveolus resident cells in ventilator-injured lungs', American Journal of Respiratory and Critical Care Medicine, 184, p.939-47.

The United States Pharmacopeial Convention (2013) 'Physical Tests / <776> Optical Microscopy'. USP 36 -NF 31. Rockville: USP, p.343-4. Veyries, M., Faurisson, F., Joly-Guillou, M. and Rouveix, B. (2000) 'Control of staphylococcal adhesion to polymethylmethacrylate and enhancement of

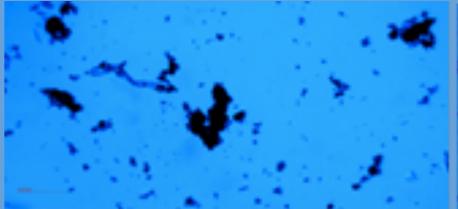
susceptibility to antibiotics by Poloxamer 407', Antimicrobial Agents and Chemotherapy, 44 (4), p.1093-96. Zabner, J., Seiler, M., Launspach, J., Karp, P., Kearney, W., Look, D., Smith, J. and Welsh, M. (2000) 'The osmolyte xylitol reduces the salt concentration of airway surface liquid and may enhance bacterial killing', Proceedings of the National Academy of Sciences of the United States of America, 97 (21), p.11614-19.

poster and further information, please scan the QR code

For a copy of this



Figure 2. Particle size distribution of Loxasperse 6.4% in purified water



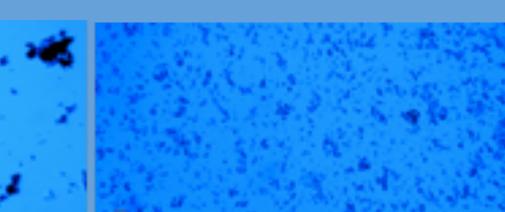


Figure 3. (left) Itraconazole 1% in purified water (right) Itraconazole 1% and Loxasperse in purified water, both at 400x magnification and blue coloured

Table 1. MIC (µg/mL) of itraconazole and itraconazole with Loxasperse (9:1) against 4 fungal strains (filamentous and yeast)

Fungal strains	A.fumigatus ATCC 204305	A.niger ATCC 16404	C.albicans ATCC 90028	R.oryzae ATCC 9363
Itraconazole	0.5	0.5	≤0.125	0.25
Itraconazole +Loxasperse	0.2	0.2	0.025	0.20

Table 2. MIC (%) of Loxasperse against 7 microbial strains

Microbial	Concentration of Loxasperse				
strains	15%	16%	17%	18%	
E.coli ATCC 8739	Growth	Growth	No growth	No growth	
S.aureus ATCC 6538	Growth	Growth	No growth	No growth	
P.aeruginosa ATCC 9027	Growth	Growth	No growth	No growth	
C.albicans ATCC 10231	Growth	Growth	No growth	No growth	
A.niger ATCC 16404	Growth	Growth	No growth	No growth	
S.typhimurium ATCC 14028	Growth	No growth	No growth	No growth	
S.aureus MRSA ATCC 33591	Growth	No growth	No growth	No growth	

Table 3. Water activity of Loxasperse, with and without desiccant after 90 days of storage at three different temperatures

Temperature	Water Activity (a _w) (with desiccant)	Water Activity (a _w) (without desiccant)
T=4°C (±1°C)	0.297	0.409
T=25°C (±1°C)	0.321	0.471
T=45°C (±1°C)	0.344	0.489